

Rootstock propagation method—How much does it matter?

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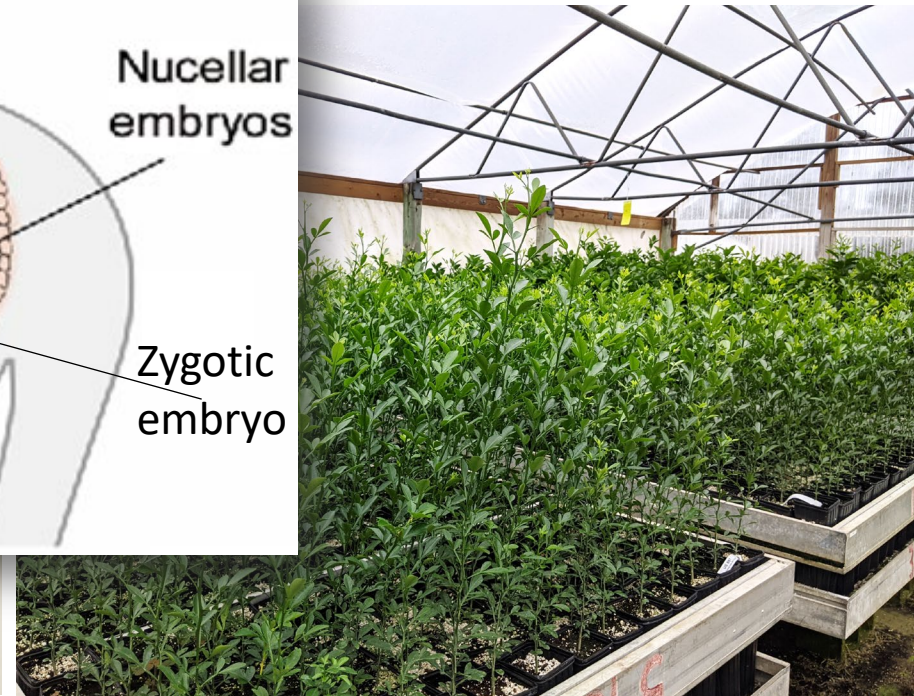
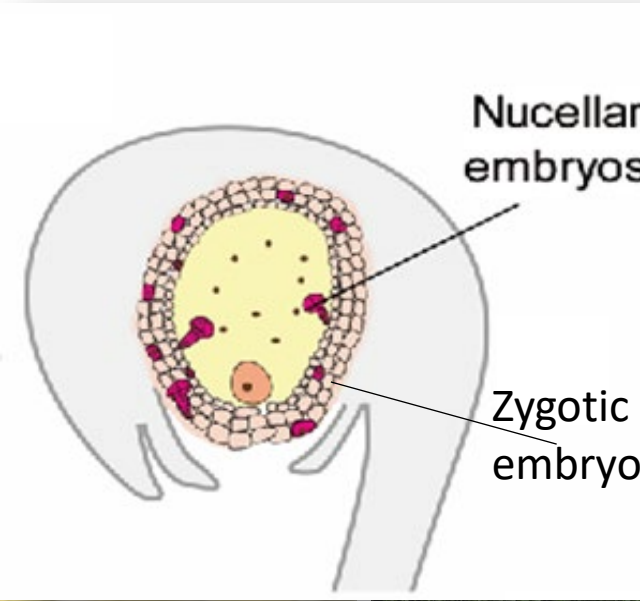


ISCN/CCNS 2022

4 Oct 2022



Seed propagation



Nucellar embryony (polyembryony)

Genetically identical embryos develop from the nucellar tissue

Cuttings and tissue culture propagation



Photo credit: Beth Lamb, Phil Rucks Nursery

Both methods produce genetically identical plants

Tissue culture propagation



Fast year-round production of uniform plants

Recent rootstock releases

Rootstock	Parentage	Released	True-to-type
US-1279	'Changsha' x 'Gotha Road'	2014	0
US-1281	'Cleopatra' x 'Gotha Road'	2014	0
US-1282	'Cleopatra' x 'Gotha Road'	2014	0
US-1283	'Ninkat' x 'Gotha Road'	2014	>90%
US-1284	'Ninkat' x 'Gotha Road'	2014	>90%
US SuperSour 1	Pummelo x 'Cleopatra'	2018	0
US Supersour 2	TF x (SO x <i>C. ichangensis</i>)	2018	TBD
US SuperSour 3	'Sunki' x US-802	2018	TBD

Top 15 Rootstocks 2020-2021

	2020	# Budded	2019	2018	2017	2016
1	US-942	1,285,560	US-942	US-942	Swingle	Kuharske
2	Kuharske	841,448	Kuharske	Swingle	US-942	X-639
3	X-639	678,095	X-639	Kuharske	X-639	
4	Swingle	468,558	Swingle	X-639	Kuharske	
5	Own Root	408,793	US-897	Sour Orange	Sour Orange	
6	US-812	296,664	US-812	US-802	US-802	
7	Sour Orange	176,322	Sour Orange	Volkamer	US-897	
8	US-897	160,288	US-802	US-812	UFR-04	
9	Volkamer	135,977	Volkamer	US-897	US-812	
10	US-802	119,887	C-54	Rough Lemon	C-35	
11	Rough Lemon	57,941	Rough Lemon	C-35	Cleopatra	
12	C-35	39,142	UFR-04	UFR-04	Volkamer	
13	C-54	26,993	C-35	UFR-17	UFR-03	
14	Poncirus trifoliata	18,106	C-57	Poncirus trifoliata	C-22	
15	UFR-04	17,892	US-1777	US-1516	Carizzo	Rough Lemon

CITRUS BUDWOOD

Annual Report
2020-2021



Seed	Tissue Culture	Rooted Cutting
38 different rootstocks used	22 different rootstocks used	17 different rootstocks used
3,550,947 propagations	650,090 propagations	208,637 propagations
Top Seed = Kuharske (790,907 Propagations)	Top Tissue Culture = US 942 (583,560 Propagations)	Top Rooted Cutting = US-942 (60,899 Propagations)

Objectives

Short-term (nursery)

- Evaluate plant traits during the nursery stage:

Long-term (field)

- Evaluate root structure, survival, and field performance during the early years and throughout the productive years.



Cuttings propagation

- Starting material: single node stem cuttings from 2-5-month-old branches
- Rooting was stimulated by applying rooting powder (Hormodine) to the basal end of the cutting
- Cuttings were maintained shaded and on an automated mist bench until acclimated



Tissue culture (TC) propagation

- Starting material: buds from mature disease-free and true-to-type plants
- Culturing on agar nutrient medium (composition proprietary)
- Sub-culturing to generate multiple shoot clusters
- Separation into individual shoots and planting in potting medium

Albrecht et al 2017. Hort Science 52(11): 1569-1576



Photo credit: Beth Lamb, Phil Rucks Nursery

NURSERY



Seedling



Root system differences

Cutting

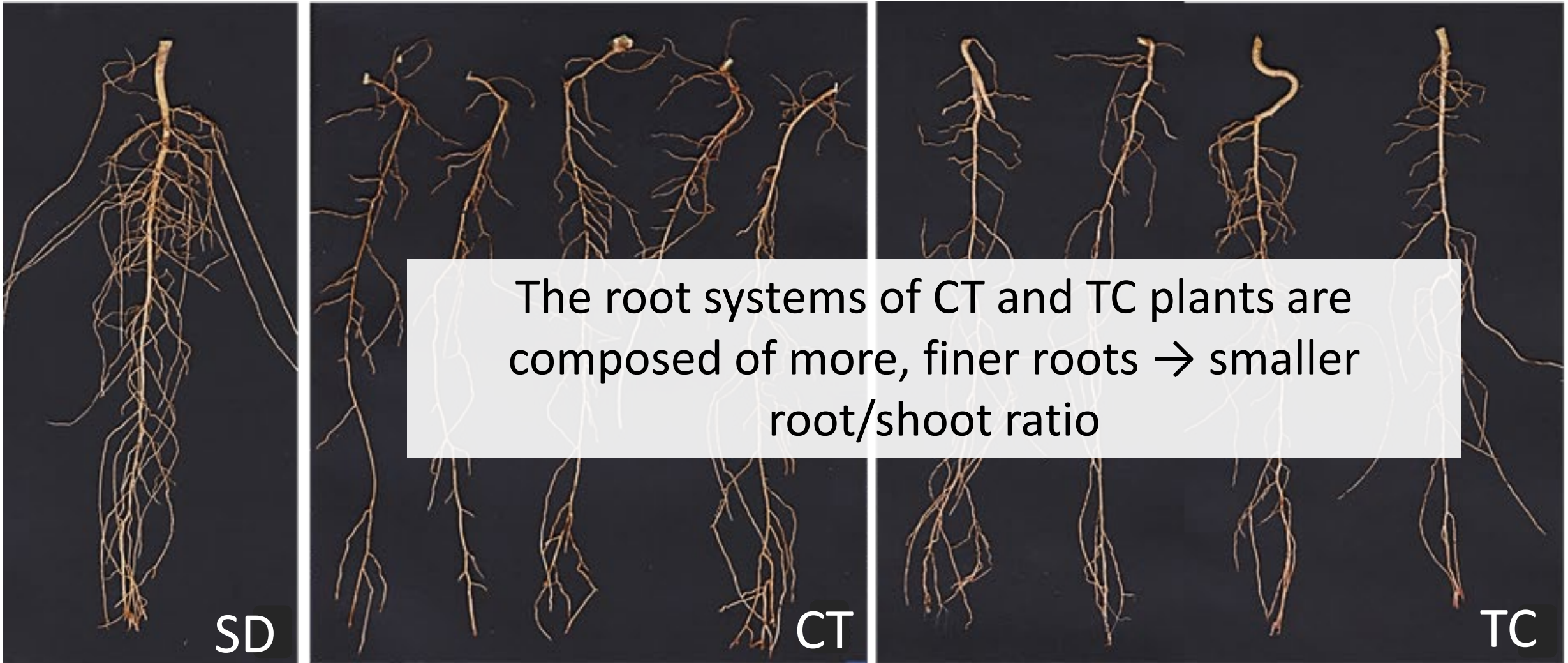


Tissue culture



Adventitious
root systems

Root system differences



Field-ready grafted trees



Root mass fractions

	Number of primary roots	Large size root mass (%)	Medium size root mass (%)	Fibrous root mass (%)
SD	1.4 c	55.8 a	20.6 b	23.6
CT	4.9 a	32.4 b	40.3 a	27.3
TC	3.6 b	42.8 b	34.2 a	23.1
	***	***	***	NS

Root system differences persist in the field-ready plants

Biomass distribution

	R/S biomass ratio	SLA (cm ² /g)	Leaf mass (%)	Stem mass (%)	Root mass (%)
SD	0.384 a	163.4	26.0	46.3 b	27.7 a
CT	0.310 b	157.8	24.7	51.8 a	23.5 b
TC	0.330 b	160.9	24.9	50.6 a	24.6 b
	***	NS	NS	***	***

Differences persist in the field-ready plants

FIELD



Airspade tree excavations

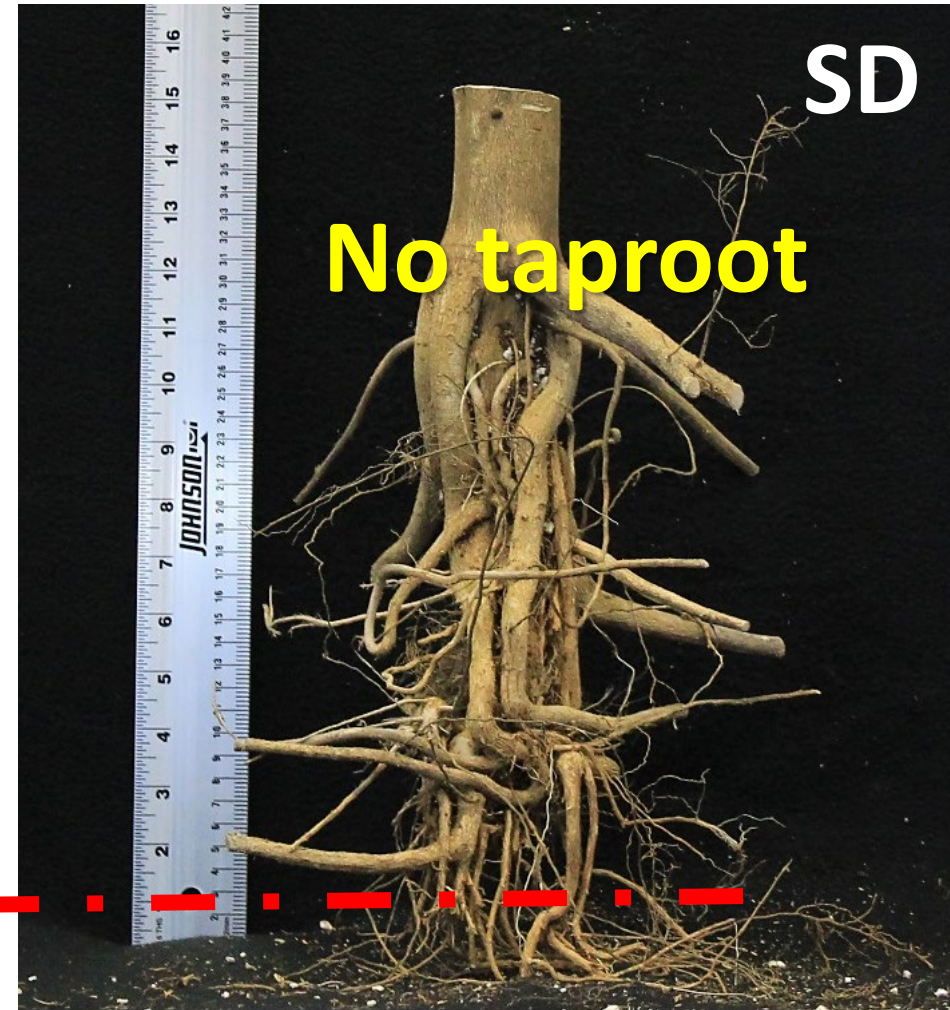
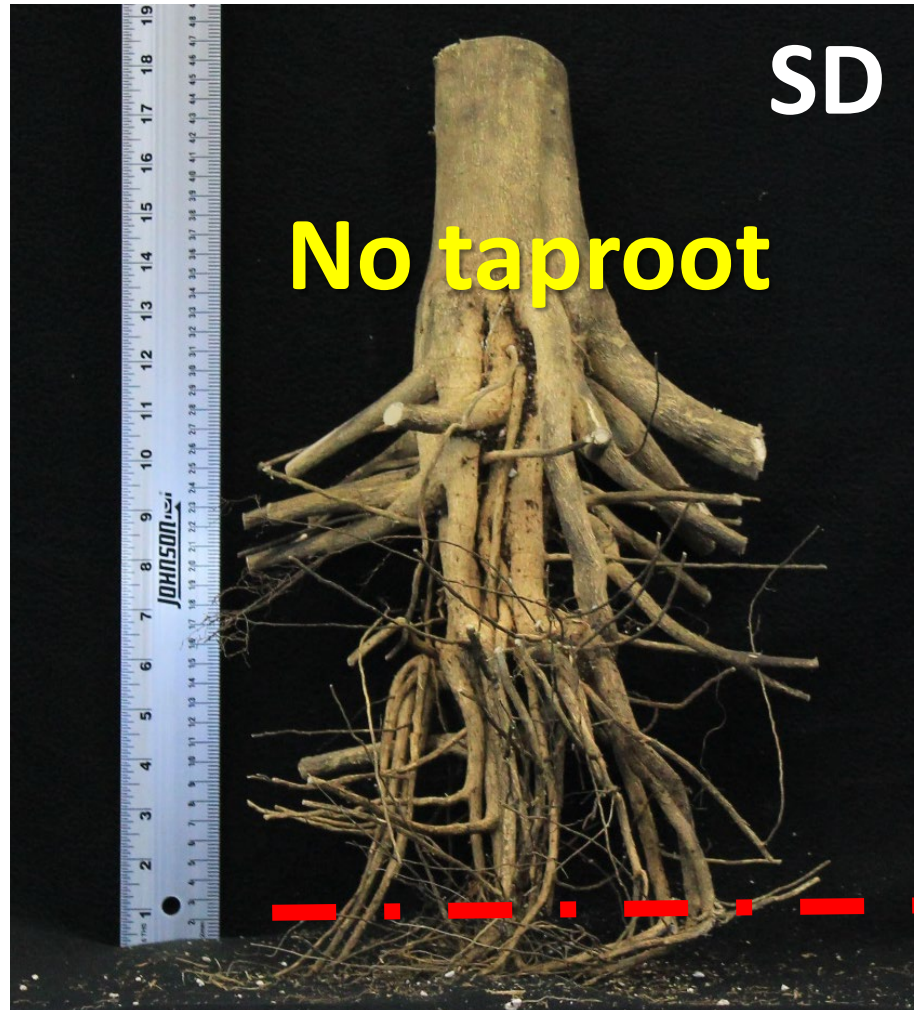


Pokhrel et al. 2020, HortScience 56 (2), 163-172

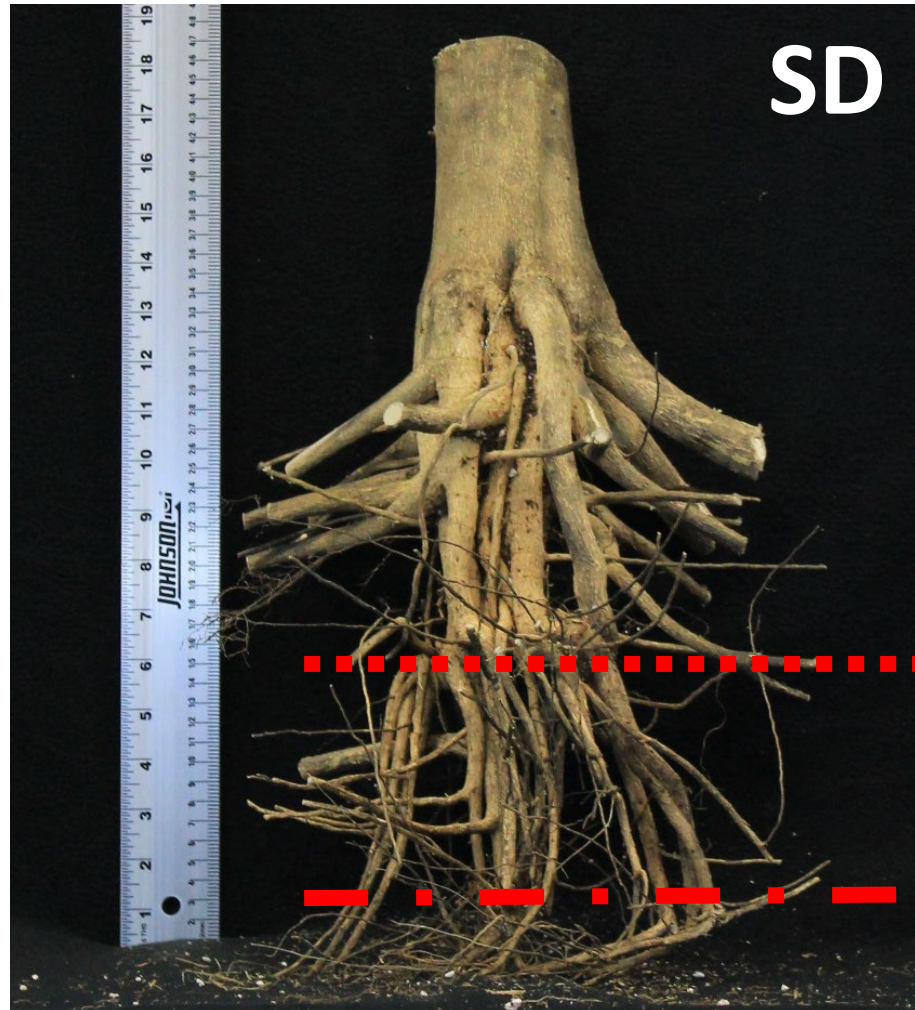


Shallow root systems—no taproot, regardless of the propagation method

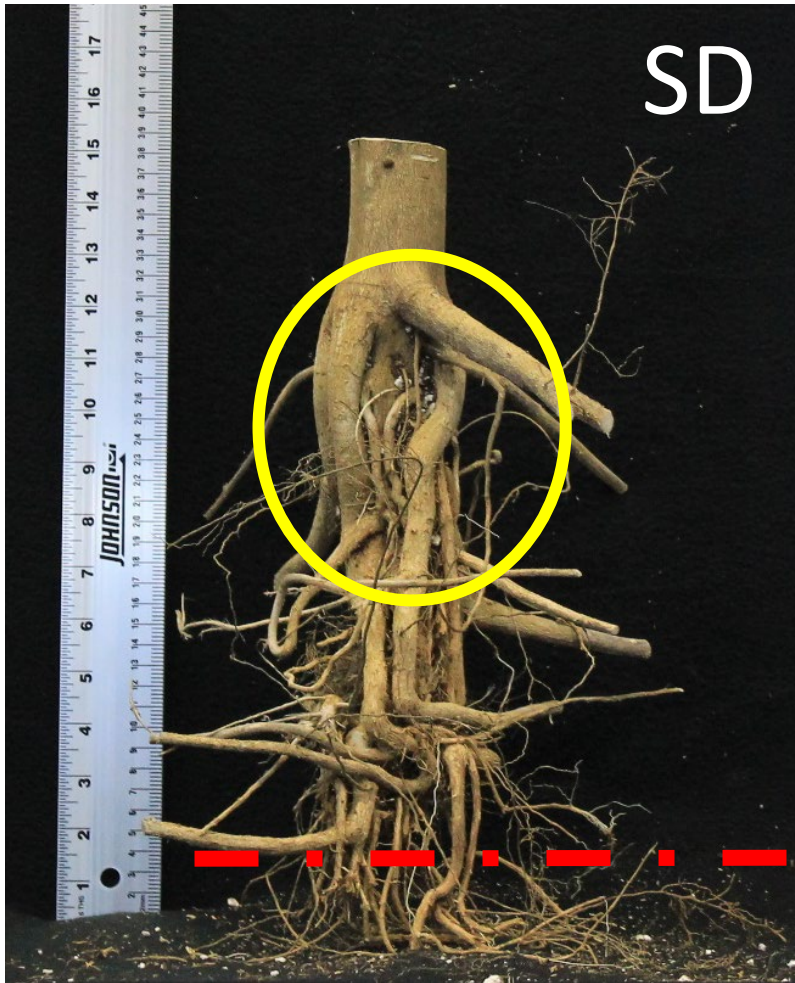
Root crown analysis



Root crown analysis

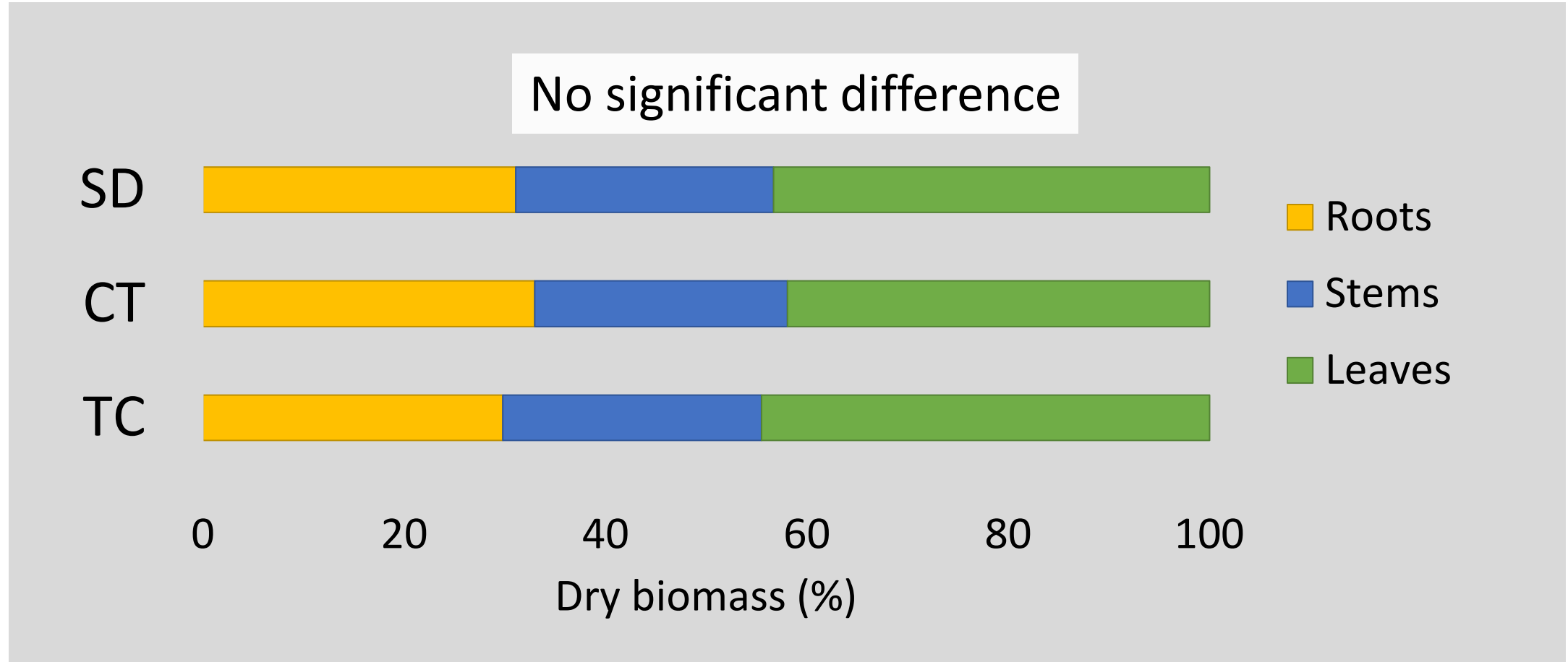


Root crowns



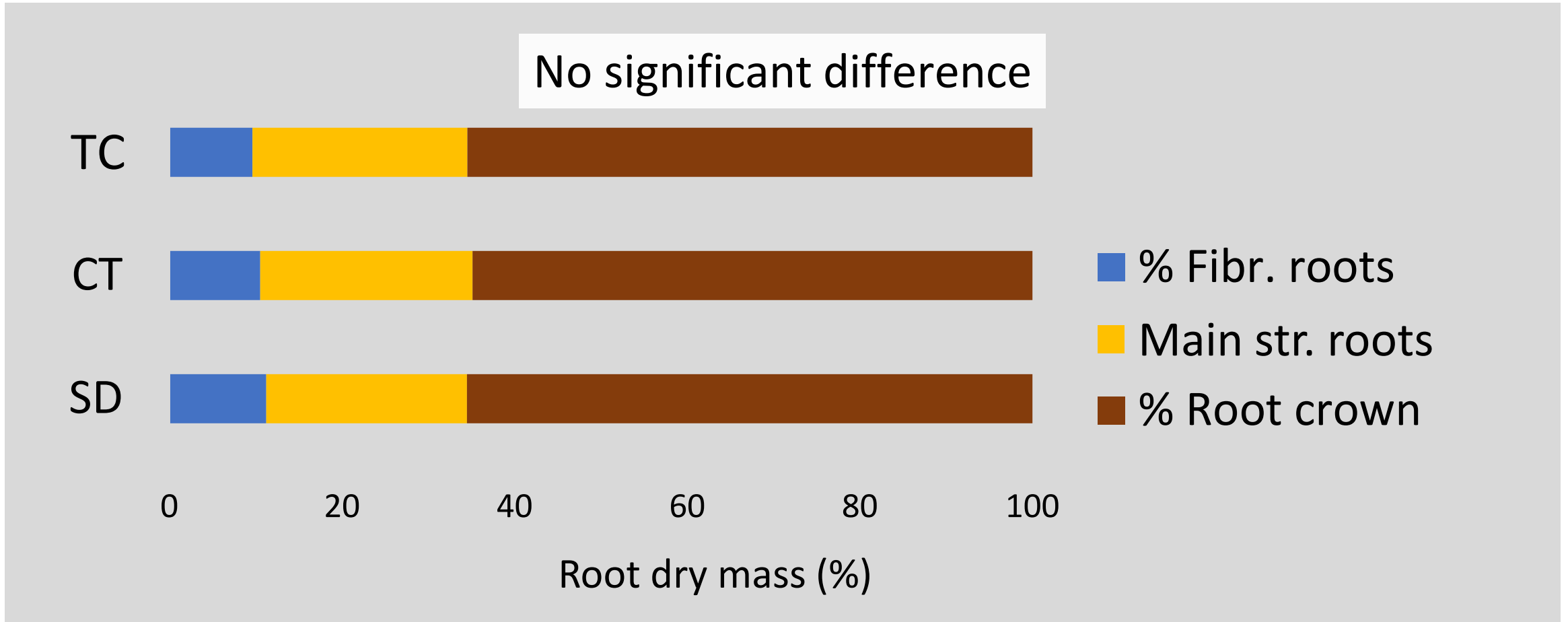
Root system differences are less noticeable after 2 years of field growth

Biomass distribution



The propagation method did not influence the tree biomass distribution

Root mass fraction

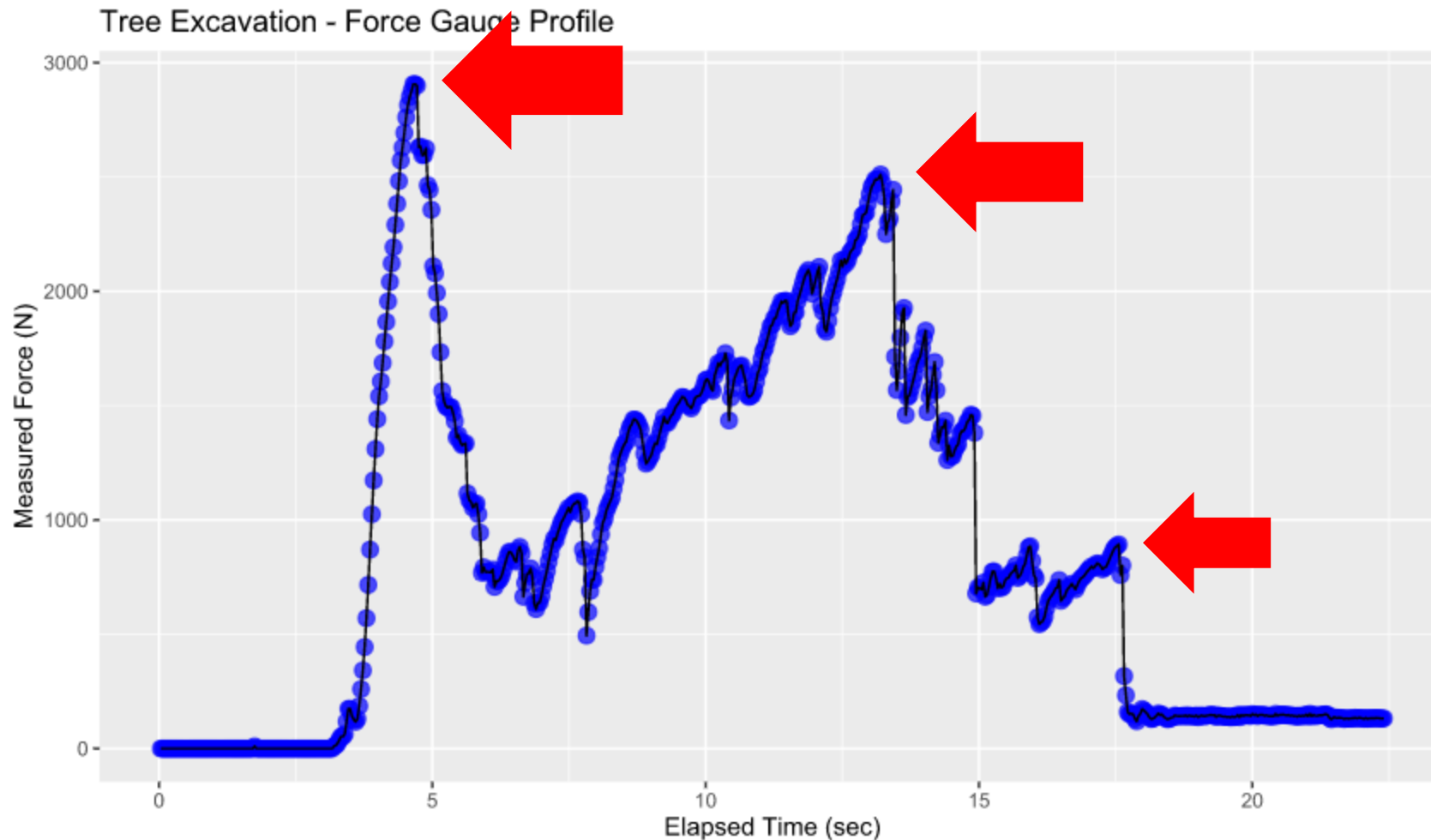


The propagation method did not influence the root mass fraction

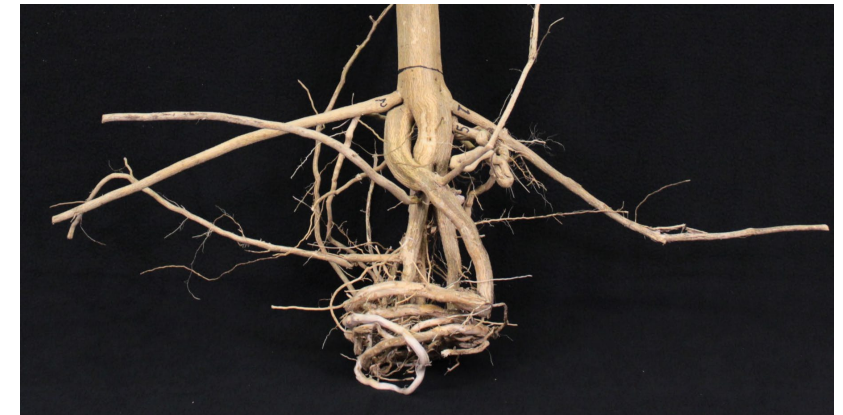
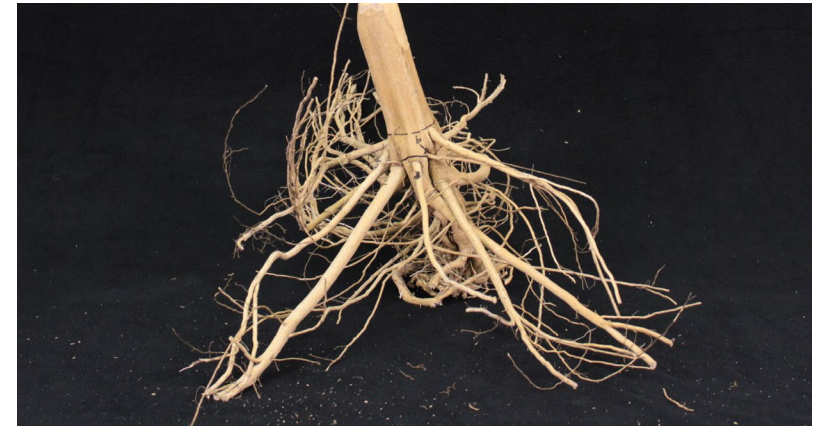
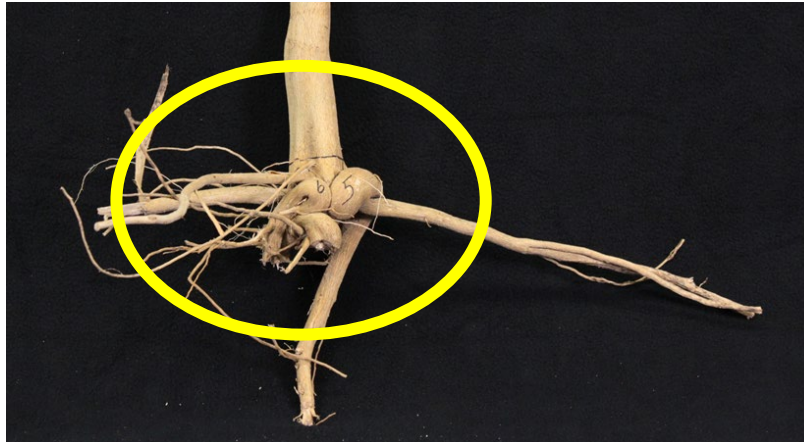
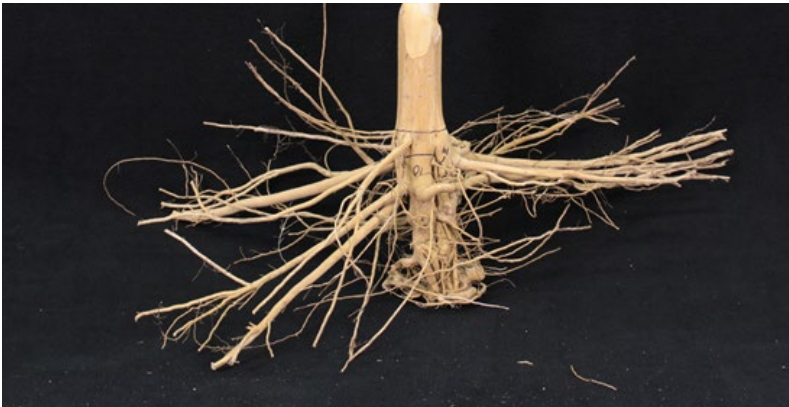
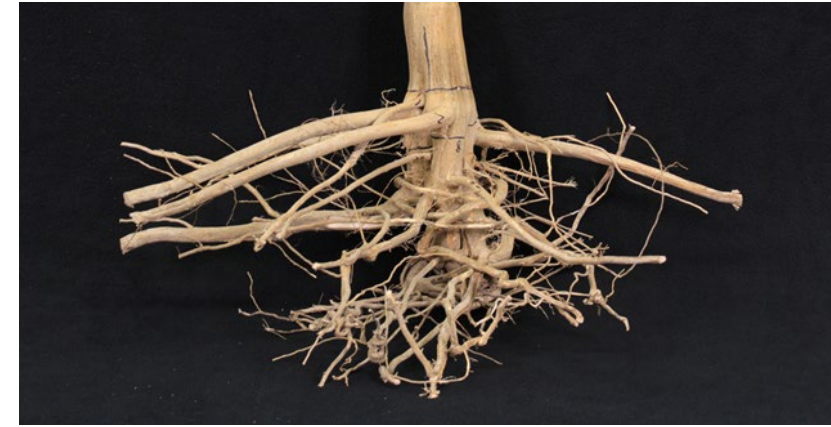
OTHER OBSERVATIONS



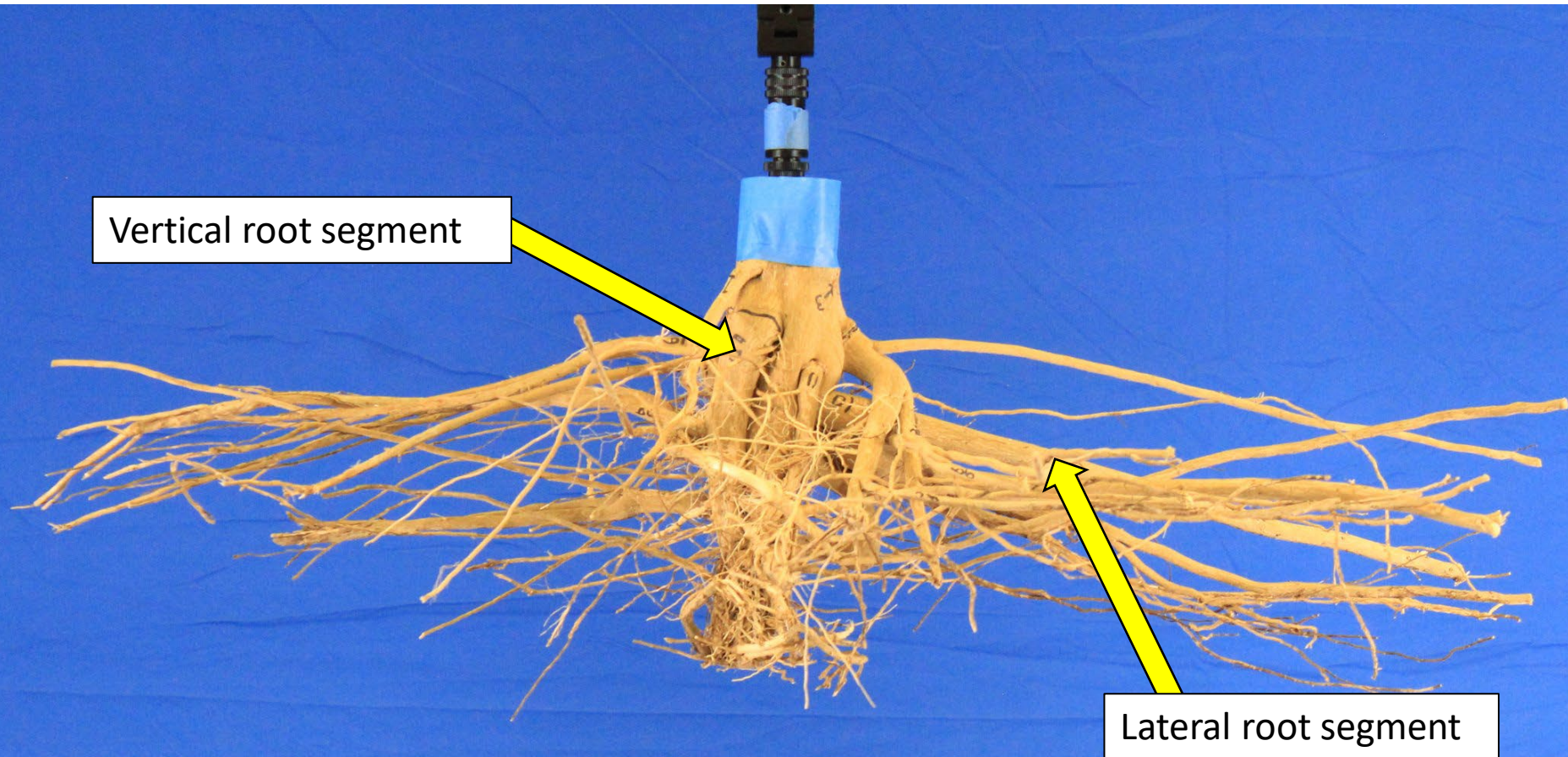
Root structure and uprooting resistance



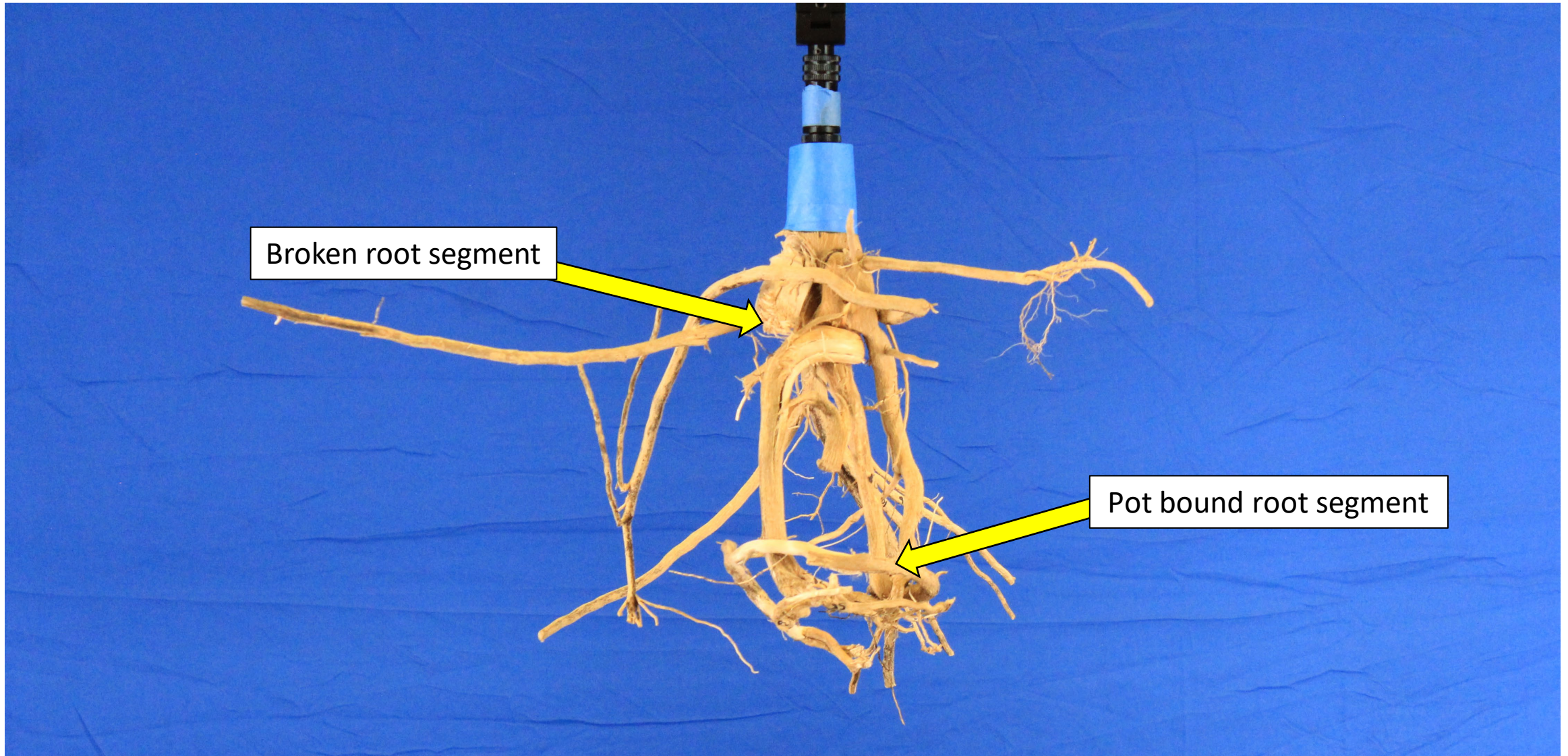
Root system phenotypes



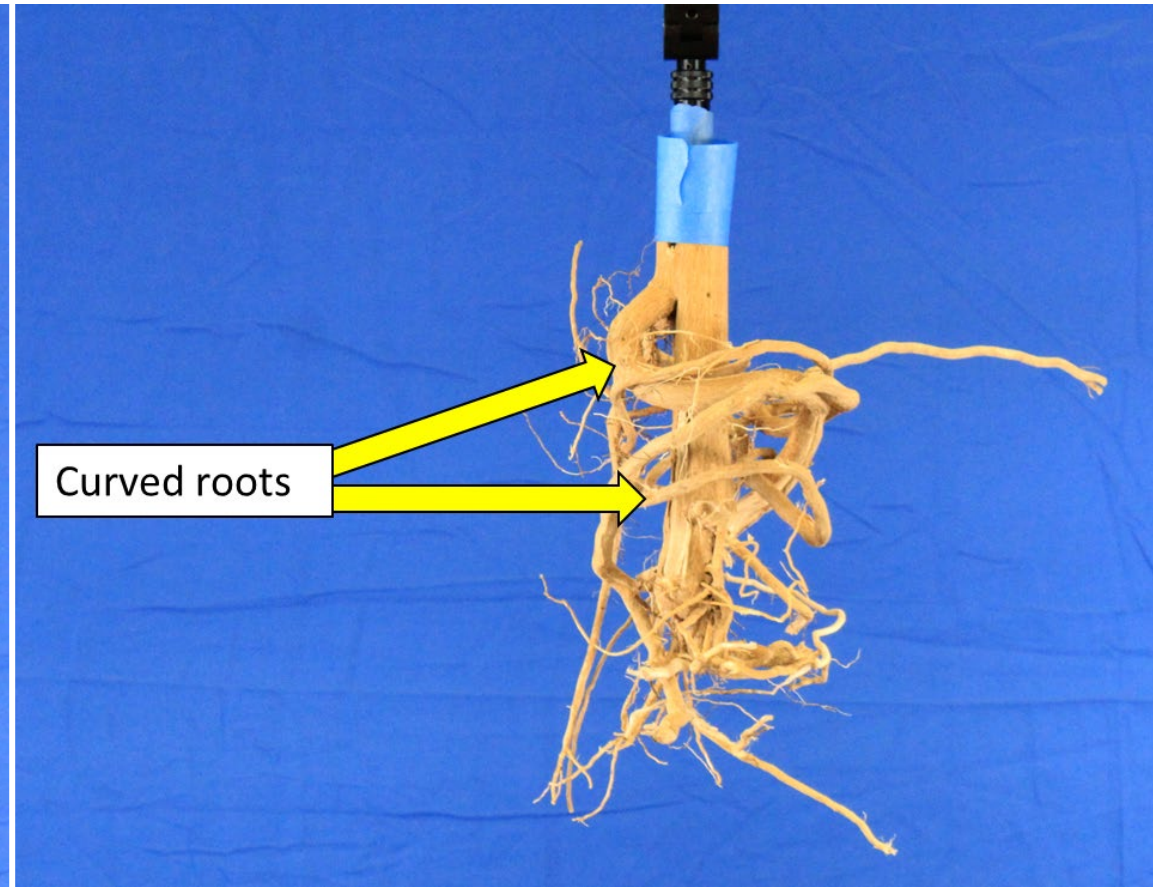
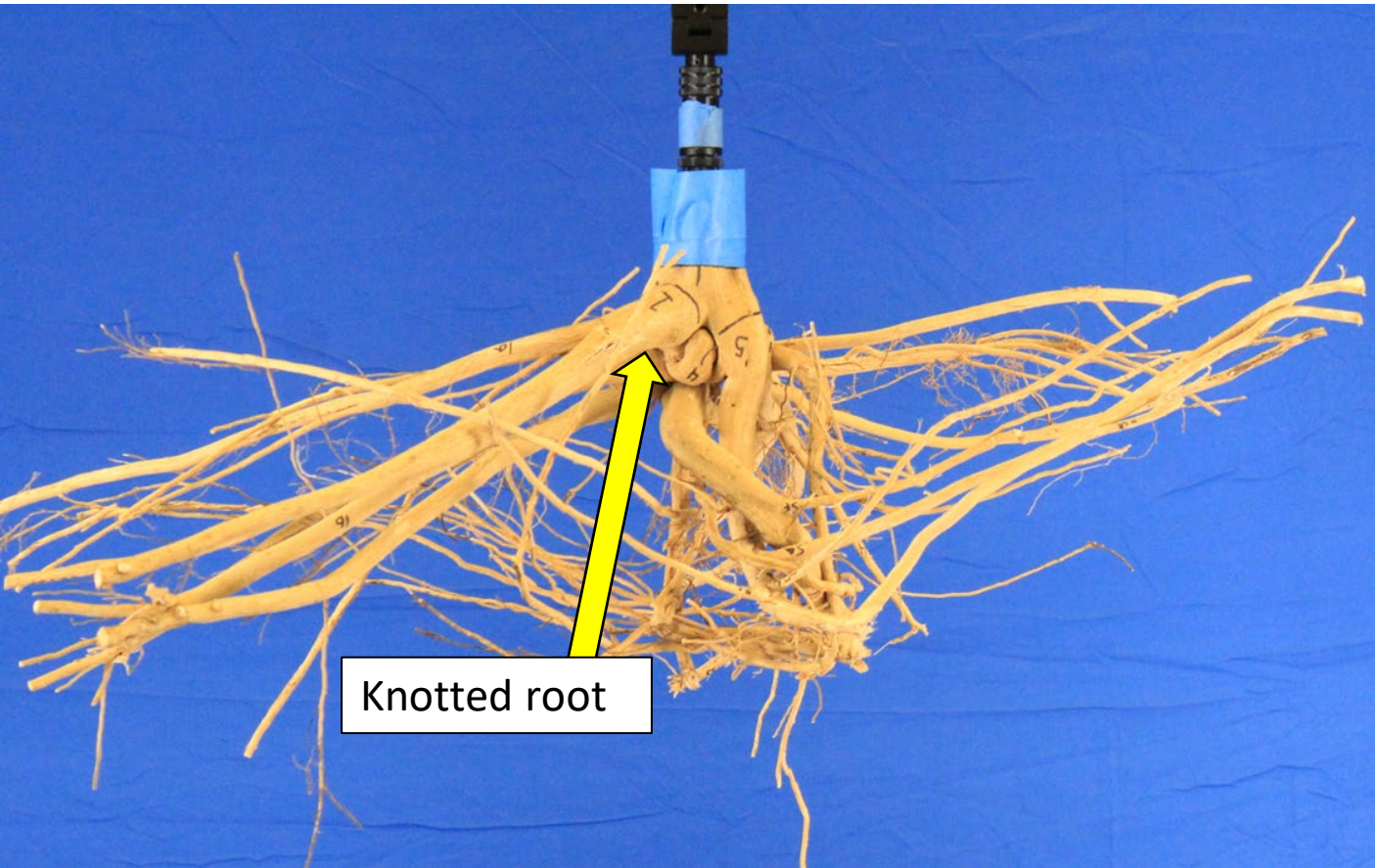
Root system analysis



Root system analysis



Root system analysis

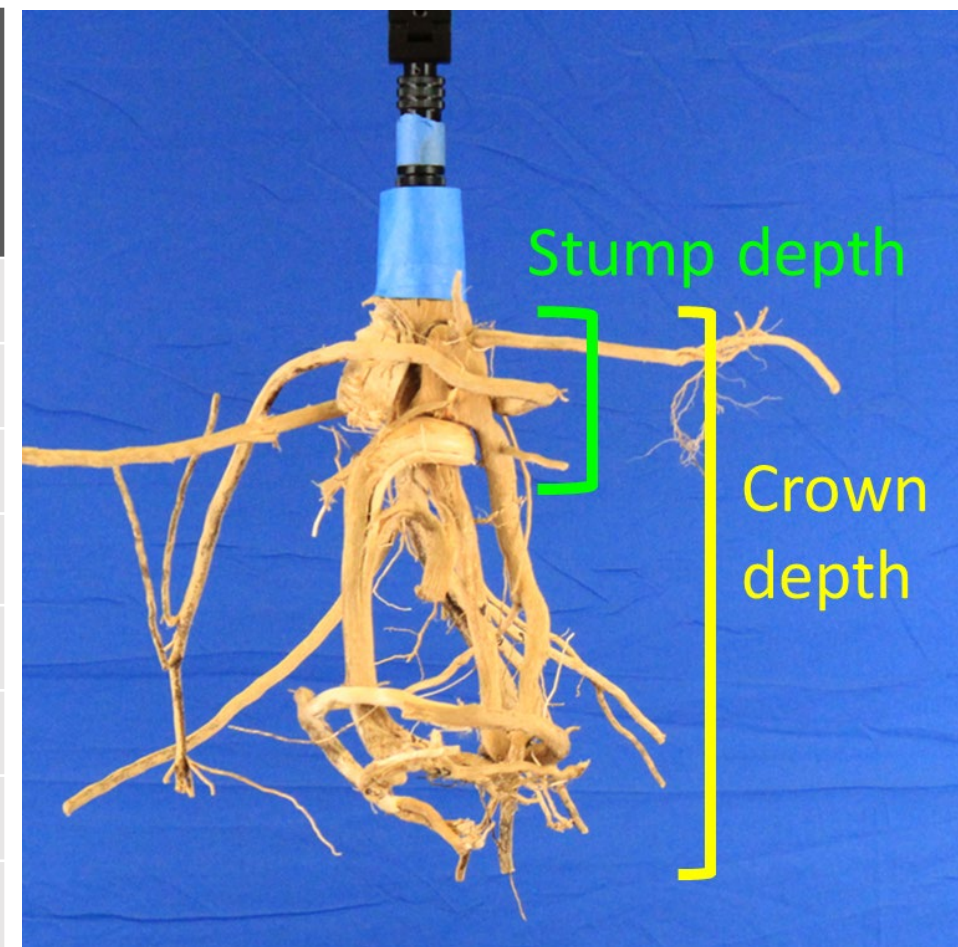


Root segments

		Lateral (%)	Vertical (%)	Straight (%)	Curved (%)	Pot-bound (%)	Knotted (%)	Broken (%)
TRIAL 1	SD	0.65 ab	0.29 ab	0.34	0.58	0.32	0.39	0.44
	CT	0.60 b	0.35 a	0.34	0.58	0.32	0.42	0.42
	TC	0.74 a	0.23 b	0.39	0.55	0.29	0.37	0.42
		*	***	NS	NS	NS	NS	NS
TRIAL 2	SD	0.68 b	0.27 a	0.29	0.62	0.28	0.42	0.42
	CT	0.71 ab	0.26 a	0.35	0.60	0.24	0.44	0.44
	TC	0.78 a	0.18 b	0.41	0.54	0.25	0.40	0.40
		*	***	NS	NS	NS	NS	NS

Root crown and stump

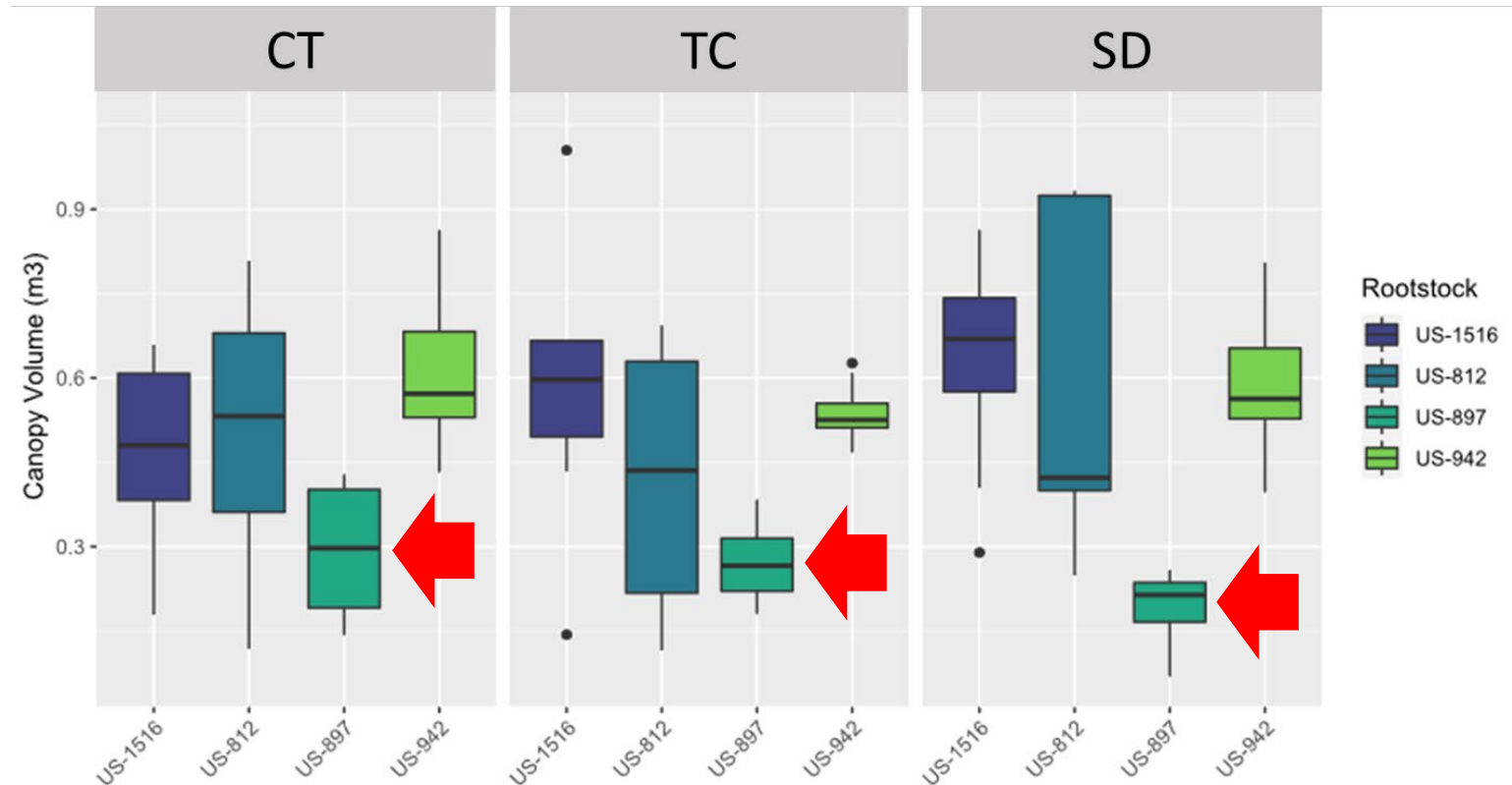
		Crown depth (cm)	Stump depth (cm)	Crown weight (g)	Stump dia (cm)
TRIAL 1	SD	26.0	22.0 a	368	3.8 b
	CT	26.1	19.6 b	321	3.8 b
	TC	25.7	9.9 c	335	4.5 a
		NS	***	NS	***
TRIAL 2	SD	23.2	20.4 a	333	4.0 b
	CT	22.7	18.9 a	369	4.1 b
	TC	22.4	10.7 b	364	4.9 a
		NS	***	NS	***





Rootstock effects and other factors

- More differences found among **rootstocks** than among propagation methods
- Other factors (planting, soil environment, etc.) influence the root structure and therefore the grafted tree performance



Thank You



UF | IFAS
UNIVERSITY of FLORIDA

Dr. Kim Bowman

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U.S. DEPARTMENT OF AGRICULTURE

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Nursery and Grower Collaborators